Thermal, morphological, and mechanical properties of ethyl vanillin immobilized in polyvinyl alcohol by electrospinning process

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Abstract In this study, polyvinyl alcohol (PVA) nanofibers with ethyl vanillin as an active compound were prepared using electrospinning technique. The final products of electrospinning process were in the form of nanofibers films. PVA/ethyl vanillin nanofibers, having fibers diameters in the range 100-1700 nm, were successfully electrospun from ethanol/water mixture of PVA and ethyl vanillin. The effects of immobilization process on ethyl vanillin thermal properties were investigated by differential scanning calorimetry (DSC). The results of DSC showed significant influence of immobilization process on thermal properties of ethyl vanillin. It was noticed that melting point of immobilized ethyl vanillin was lower (~55 °C) compared to free flavor (\sim 77 °C). Our results showed that films based on PVA/ethyl vanillin nanofibers are mechanically stable.

Keywords Polyvinyl alcohol · Ethyl vanillin · Electrospinning · DSC · Nanofibers

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Introduction

The productions of polymer films that contain active compounds by electrospinning process have attracted great attention in past decades. Production of films by electrospinning process is based on polymers properties that allow the formation of nano-scale fibers under electrostatic force. The formation of nanofibers is influenced by several parameters: intensity of applied electrical potential, polymer characteristics (i.e., viscosity, conductivity and surface tension), flow rate of polymer solution, and distance between two charged electrodes (i.e., needle and collector). Generally, the electrospinning set-up (Fig. 1) is consisting of dosage system (usually syringe pump with syringe and blunt metal needle), collector and high voltage dc generator.

Electrostatic field causes polymer jet formation at the end of a needle tip, which is attracted by opposite charged collector. Liquid jet formation occurs, when electrostatic force overcomes the surface tension of the polymer solution and polymer solution at the needle tip forms conical shape—Taylor cone. During the jet transition from needle to collector, the fibers are formed simultaneously with a solvent evaporation. As a results, the film consisted of nanofibers is formed on collector's surface [1–3].

Polyvinyl alcohol (PVA) is a non-toxic synthetic polymer that has been used in food industry as moisture barrier, or as an active packaging material. Also, in some products, PVA can be applied as a coating material against negative influence of atmospheric oxygen [4]. PVA is commonly used for production of nanofibers by electrospinning process. These fibers have been designed as carriers for immobilization of various materials like a drugs or food additive [5, 6]. The morphological properties of electrospun PVA nanofibers can be regulated by the addition of



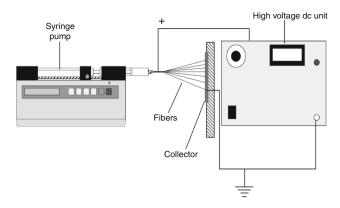


Fig. 1 Schematic of electrospinning process

surfactant in polymer solution prior to electrospinning process [7] or using inorganic materials [8].

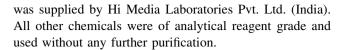
Ethyl vanillin (3-ethoxy-4-hydroxybenzaldehyde) has been used as a flavor in food products and cosmetics. This synthetic flavor has more intensive flavoring power compared to vanillin (3-methoxy 4-hydroxybenzaldehyde). On the other hand, vanillin is more soluble in some solvents and solvents mixtures than ethyl vanillin. The solubility of ethyl vanillin in water is lower compared to vanillin, while it is soluble in organic solvents (i.e., ethanol) [9, 10]. Recently, electrospinning technique has been successfully applied for immobilization of the vanillin in PVA and cyclodextrin. This approach showed significant potential of electrospinning as method for production of immobilized flavor with improved thermal stability and prolonged shelf-life [6].

The goal of this study was to investigate the production of PVA films that contain ethyl vanillin as an active flavor compound. Electrospinning technique was applied for production of nanofibrous films. Since ethyl vanillin is practically insoluble in water, the mixture of ethanol/water was used as a solvent system for dissolving of flavor. The morphology of nanofibers was characterized by scanning electron microscopy (SEM). The effects of the immobilized flavor, as well as ethanol/water solvent system on the morphology of PVA nanofiber were evaluated. The free flavor, PVA film and PVA/ ethyl vanillin nanofibrous films were characterized by Fourier transform infrared spectroscopy (FTIR), X-ray diffraction analysis, differential scanning calorimetry (DSC), and thermogravimetry (TG). In order to evaluate the influences of immobilized flavor on the mechanical properties of electrospun films, the tensile stress of samples was analyzed.

Experimental

Materials

PVA (average Mw = 70.000-100.000, hot water-soluble) was purchased from Sigma-Aldrich (USA). Ethyl vanillin



Preparation of the flavor and polymer solutions

In this study, ethanol—water (volume ratio ethanol—water = 70:30) mixture was used as a solvent for ethyl vanillin. The working solution (20 % m/v) of ethyl vanillin was prepared by dissolving of flavor in ethanol—water mixture.

PVA (0.15 g mL⁻¹) was dissolved in distillated water at 80 °C. Aqueous solution of PVA was mixed with ethanol—water and ethanol—water/ethyl vanillin solution (See Table 1) and stirred on magnetic stirrer for 15 min. The final volume of all mixtures was 3 mL. In addition, 2 mL of ethanol—water solution was added in order to provide complete flavor dissolution.

Electrospinning procedure

The syringe (5 mL) with blunt stainless still needle (18 G) was fixed horizontally on the syringe pump (Pump 11, Harvard Apparatus, US) and the solutions were electrospun by high voltage power supply (Model 30R, Bertan Associates, US). The other process parameters were: flow rate-0.3 mL h⁻¹, applied voltage-13 kV, and needle tip to collector distance-10 cm. A grounded piece of aluminum foil was used for the fiber deposition. The formed films were dried at room temperature for 24 h to remove the residual solvents mixture if any present.

Measurement and characterization

The diameters and morphology of the nanofibers were investigated by SEM (JEOL JSM-6390LV). The samples were coated with gold on BALTEC SCD 005 sputter coater prior to SEM analysis. The diameter of fibers was analyzed using ImageJ image analysis program. Around 100 fibers from the SEM images were used for diameter analysis. The thickness of nanofibrous films was measured by a micrometer caliper.

DSC measurements were performed in a DSC-50 (SHIMADZU) DSC thermal analyzer using sample mass of 10.0 mg at heating rates of 10 °C min⁻¹ under air atmosphere.

Thermogravimetric analysis was carried out by a SETARAM SETSYS Evolution-1750 instrument. The measurements were conducted at a heating rate of 10 °C min⁻¹ in air atmosphere. FTIR spectrum of free ethyl vanillin, PVA film, and PVA film with flavor was studied using Bomem-Hartmann & Braun (USA) spectrometer. Spectra were collected using KBr pellets in the spectral range 4,000–600 cm⁻¹, with the resolution of 4 cm⁻¹.



Table 1 Compositions of solution used in experiment and fibers/films characteristics

Solution composition	PVA/ethanol- water/mass ratio	Ethyl vanillin concentration/%w/v in ethanol–water solution	Fibers characteristics	Tensile stress/MPa
PVA solution	100/0	-	None ^b	_
PVA solution/ethanol-water	70/30 ^a	-	Fibers with beads/diameters in the range 100–800 nm	8.4 ± 2.8
PVA solution/ethanol-water/ethyl vanillin	70/30 ^a	20	Fibers with beads/diameters in the range 100–1700 nm	23.3 ± 4.3

^a Additionally add 2 mL of ethanol-water solution

The X-ray powder diffraction measurements were performed on a Philips 1050 X-ray powder diffractometer using Ni-filtered Cu Ka radiation and Bragg–Brentano focusing geometry. The patterns were taken in the 5–70° 2θ range with the step of 0.05° and exposure time of 6 s per step.

The mechanical properties of the PVA and PVA/ethyl vanillin films were analyzed using an Autograph AG–X Plus High Speed Universal Testing Machine (SHIMADZU). All of the tensile measurements were carried out in triplicate, at room temperature, while the test speed was kept at 1.0 mm min^{-1} . The dimensions of the sheet used were $\sim 1 \times 2 \text{ cm} \times \sim 30\text{-}60 \text{ }\mu\text{m}$ (width \times length \times thickness).

Results and discussion

Effects of ethyl vanillin on morphological properties of nanofibrous films

The obtained PVA films were colorless and transparent, while those with immobilized flavor were nontransparent and white. The thickness of PVA films was found to vary in the range of $\sim 30~\mu m$ (for PVA films) to $\sim 60~\mu m$ (for PVA films with immobilized flavor).

Effects of ethyl vanillin addition on fibers formation and fibers diameters

The morphological properties, as well as fibers diameters of the electrospun films were examined using the SEM. The diameter values and images of electrospun PVA and PVA/ethyl vanillin fibers are shown in Fig. 2. The diameters of PVA nanofibers vary from 100 to 800 nm. It can be seen from Fig. 2a, the PVA fibers are irregular and interspersed with beads. These nanofibers properties are in good agreement with results previously reported by Zhang et al. [11] for electrospinning of PVA with the addition of ethanol. According to the same authors,

ethanol is not solvent for PVA and causes the reduction of polymer surface tension, as well as viscosity. Hence, as a result of the electrospinning process of PVA/ethanol mixtures, the fibers with beads were formed. On the other hand, the usage of ethanol as a solvent in this study is necessary due to low solubility of ethyl vanillin in water.

The SEM images and diameter distribution histogram for the electrospun PVA film with immobilized ethyl vanillin are shown in Fig. 2b. It can be noticed that for PVA/ethyl vanillin film, fibers diameters are in the range 100–1,700 nm, while the addition of flavor changes not only the morphology of fibers, but also increases the fibers diameters.

Mechanical properties of nanofibrous PVA/ethyl vanillin films

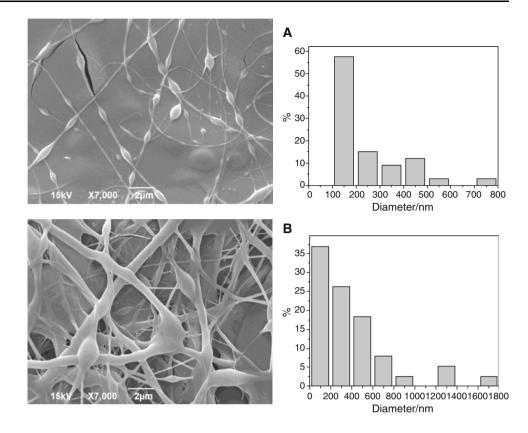
The effects of immobilized ethyl vanillin on mechanical properties of PVA nanofibrous films are studied and results are shown in Table 1. Our results showed that immobilized flavor affects significantly mechanical properties of PVA nanofibrous film. The tensile stress increased from 8.4 ± 2.8 for PVA films to 23.3 ± 4.3 MPa for PVA/ethyl vanillin films. As we showed above, immobilized ethyl vanillin acts as filler substance and causes increasing of fibers diameters and changed the fiber morphology. As we noticed above, the immobilization of flavor changed the morphology of nanofibers. Hence, variations of mechanical properties of nanofibrous PVA films with immobilized flavor can be expected.

Mollá et al. [12] showed that tensile strength of electrospun Nafion/PVA membranes increased with the increasing of the thickness of these membranes. Our results pointed out that tensile strength of PVA and PVA/ethyl vanillin films depends on polymer concentration, addition of ethanol and ethyl vanilline and modified conditions for fiber formation.



^b Solution was too viscous

Fig. 2 Morphology and fibers diameter distribution histograms for the electrospun PVA film without ethyl vanillin *A* and film with immobilized flavor *B*



FTIR analysis of nanofibrous PVA/ethyl vanillin films

FTIR analysis was performed in order to examine the functional groups, as well as the existence of PVA and ethyl vanillin in the electrospun nanofibers. As shown in Fig. 3, the FTIR spectrum of free ethyl vanillin displays a number of strong absorption peaks which were previously explained by Levic et al. [13]. The characteristic vibrations detected for PVA nanofibers are: at around 3,423 cm⁻¹ from OH groups; region 3,000–2,840 cm⁻¹ from CH₂ and C–H; at around 1,640 cm⁻¹ from OH bending; at around 1,430 cm⁻¹ from CH₂; 1,091 cm⁻¹ from C–O and at around 850 cm⁻¹ from C–C vibrations [14–16].

The FTIR spectrum of PVA/ethyl vanillin film, in the region between 1,750 and 600 cm⁻¹, shows clearly domination of the flavor's bands. On the other hand, in the region 4,000–2,750, the picks corresponding to OH groups, CH₂, and C–H in PVA can be clearly distinguished in the film's FTIR spectra.

X-ray powder diffraction analysis of nanofibrous PVA/ ethyl vanillin films

X-ray diffraction patterns of free ethyl vanillin, PVA nanofibrous film, and immobilized flavor are shown in Fig. 4. In the diffractogram of free ethyl vanillin, the characteristic peaks are at diffraction angles 2θ of 12.2° ,

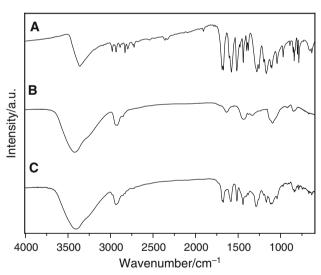


Fig. 3 FTIR spectra of free ethyl vanillin A, PVA nanofibrous films B and PVA/ethyl vanillin nanofibrous films C

14.5°, 21.1°, 22.7°, 26.4°, 29.2°, 37.45°, and 38.85°. The represented X-ray diffraction spectra of free flavor indicate crystalline structure of ethyl vanillin, and it is in accordance with previously published data [17]. The diffractogram of PVA film exhibits broad diffraction pattern at diffraction angles 2θ of 19.6° and it is characteristic for semi-crystalline nature of this type of polymer [6].



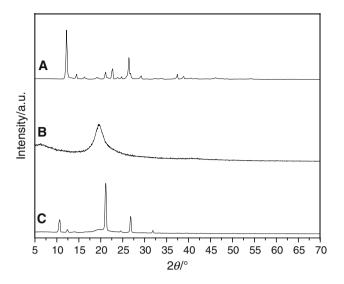


Fig. 4 X-ray diffraction patterns of the free ethyl vanillin A, PVA nanofibrous film B and PVA/ethyl vanillin nanofibrous films C

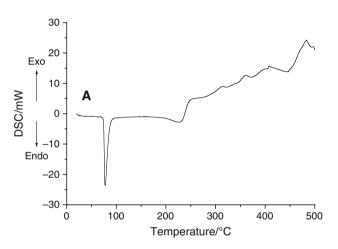


Fig. 5 DSC curve of the free ethyl vanillin A

Diffraction pattern of immobilized flavor shows peaks at diffraction angles 2θ of 10.65° , 12.35° , 21.1° , 24.55° , 26.8° , and 31.9° . The differences in the diffraction pattern of immobilized ethyl vanillin compared to free aroma indicates the modification of the flavor crystal properties which occur as a result of immobilization in PVA matrix. These modifications are most probably the result of the crystals formation, which occur due to change in electrospinning process parameters (e.g., fast solvent evaporation and presence of PVA). Our results point out that immobilized flavor retained crystalline structure after immobilization in PVA matrix. Kayaci and Uyar [6] showed that vanillin immobilized in PVA nanofibers did not show crystalline structure compared to free flavor. Similar results were reported for caffeine and riboflavin

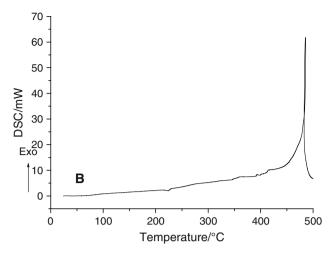


Fig. 6 DSC curve of the PVA nanofibrous films B

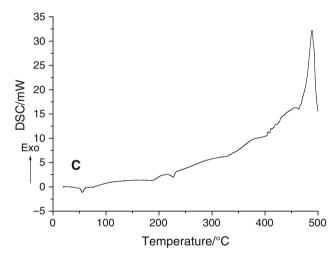


Fig. 7 DSC curve of the PVA/ethyl vanillin nanofibrous films C

immobilization into PVA nanofibers by electrospinning process [18]. The differences in our result regarding previous studies could be explained by the fact that ethyl vanillin is low soluble in water, but shows high solubility in ethanol. Ethanol primarily evaporates from water/ethanol mixture during electrospinning process as result of lower boiling point than water. This probably leads to fast flavor crystal growth and formation of crystalline aggregates in the remaining aqueous phase.

DSC analysis of nanofibrous PVA/ethyl vanillin films

Thermal analysis was performed in order to investigate thermal properties of films produced using electrospinning techniques. DSC curves of the free ethyl vanillin, PVA, and



PVA/ethyl vanillin fibers are shown in Figs. 5–7, and results of thermal analysis are summarized in Table 2. Free ethyl vanillin exhibited a sharp endothermic peak at 77.5 °C (Fig. 5), which indicates the melting point (T_m) of flavor [9]. The melting point of immobilized ethyl vanillin (Fig. 7) is shifted to lower temperature (54.9 °C). The values of melting enthalpy $(\Delta H_{\rm m})$ also decreases for immobilized ethyl vanillin compared to free flavor. The decreasing of melting point temperature has been observed for nanofibrous polymer systems [19]. Arinstein et al. [20] showed that depression of melting temperature for polymers occurs by rapidly solvent evaporation during electrospinnig process, which causes the significant influences on polymer chain organization. The depression of melting point for free ethyl vanillin might be explained by changed conditions for flavor crystallization during formation of nanofibers (See above). Also, the presence of PVA affects on thermal properties of ethyl vanillin and indicates that polymer and flavor probably create separated phases in the nanofibers. As we showed above, FTIR analysis did not indicate the presence of strong interactions between this two components.

On the other hand, in the DSC curve of PVA film (Fig. 6), the peak at 227 °C can be observed, which corresponds with melting of PVA. This peak in the case of PVA/ ethyl vanillin nanofibrous film is at ~ 226 °C. It seems that addition of ethyl vanillin did not affect significantly on PVA nanofibers melting point. The reason for this might be found in the fact that the free ethyl vanillin does not show peaks (endothermic or exothermic) in around the PVA melting point. Also, based on literature data, the bulk of ethyl vanillin evaporates before the PVA melting point is reached. According to Levic et al. [13], the main mass loss of free ethyl vanillin occurred up to 230 °C, while the encapsulated ethyl vanillin, according to the same authors showed resistance toward mass loss during thermal analysis. Taepaiboon et al. [5] showed that different active compounds may affect on thermal stability of PVA nanofiber by changing kinetics of thermal degradation of polymer. Nevertheless, for application of PVA/flavor nanofibrous film in different areas of food technology and food packaging, it is important to achieve thermal stability of system polymer/flavor in the specific temperature ranges. The applied temperature range in this study corresponds with temperature range applied in food processes [21, 22].

The temperature range from 250 to 500 °C corresponds to thermal decomposition of PVA and ethyl vanillin. The temperature maximum and corresponding enthalpies are represented as temperature (T_d) and enthalpy (ΔH_d) of decomposition. Karathanos et al. [23] showed that oxidation of vanillin is a process that most likely consists of formation of vanillic acid, while the thermal decomposition occurred at higher temperature (above 300 °C). Our result indicates that above 250 °C, the thermal decomposition of free ethyl vanillin is complex process, and probably includes both oxidation and decomposition of flavor. In the temperature range from 250 to 500 °C, the DSC curve of PVA film (Fig. 6) shows exothermic peak at 485.3 °C. According to Modafferi et al. [24], the exothermic reaction in this temperature range is attributed to the decomposition of PVA. The thermal decomposition of PVA is complex process that includes the breakdown of the polymer backbone at high temperatures. On the other hand, the DSC curve of PVA/ethyl vanillin film shows broad peak at 488.8 °C. The exothermic peak for PVA/ethyl vanillin film is occurred at almost the same temperature as in the case of PVA film. The addition of ethyl vanillin causes decreases in enthalpy compared to PVA film. Asran et al. [25] showed that addition of other compound changes the crystal structure of PVA fibers, which is observed by changes in the enthalpies.

Thermogravimetric analysis of nanofibrous PVA/ethyl vanillin films

The results of TG analyses of free ethyl vanillin, PVA nanofibrous film, and PVA/ethyl vanillin nanofibrous film are shown in Fig. 8. Free ethyl vanillin exhibits main mass loss in the temperature region 150–250 °C. The PVA film shows two step mass losses: up to 225 °C the mass loss is ~ 10 % and most probably, it is related with water release; the second characteristic mass loss occurs from 225 up to 500 °C, and it is

Table 2 DSC data obtained by analyzing the free ethyl vanillin, PVA nanofibrous films, and PVA/ethyl vanillin nanofibrous films

Sample	Temperature range from RT to 250 $^{\circ}\text{C}$		Temperature range from 250 to 500 °C	
	T _m /°C	$\Delta H_{\rm m}/{\rm kJ~g^{-1}}$	$T_{\rm d}$ /°C	$\Delta H_{\rm d}/{\rm kJ~g^{-1}}$
Free ethyl vanillin	77.54	-10.00	483.51	0.22
PVA nanofibrous films	_	_	485.31	1.05
PVA/ethyl vanillin nanofibrous films	54.95 ^a	-0.0154	488.78	0.68
	226.63 ^b	-0.0224		

^a Indicate first peak on DSC curve

b Indicate second peak on DSC curve



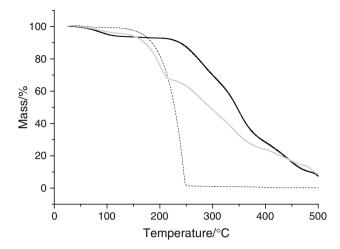


Fig. 8 TG curves of the free ethyl vanillin (*dashed line*), PVA nanofibrous film (*dark line*), and PVA/ethyl vanillin nanofibrous films (*gray line*)

associated with polymer decompositions. The PVA/ethyl vanillin TG curve exhibits more complex mass loss kinetics regarding free flavor or PVA film. The mass loss of immobilized flavor could be separated in three regions: (i) water loss from PVA film, (ii) flavor release, and (iii) finally polymer and flavor thermal decomposition. Water release occurs in the same temperature region in which ethyl vanillin starts to evaporate, so it is relatively complex to distinguish these two processes. Nevertheless, the influence of electrospinning process on ethyl vanillin thermal properties is pronounced. Up to 250 °C, the immobilized flavor shows ~ 37 % mass loss, compared to free flavor (i.e., 98 % mass loss) or PVA film (i.e., ~ 10 % mass loss).

Our results point out that electrospinning of ethyl vanillin in the form of nanofibrous film is based on PVA as matrix material modifies flavor thermal properties and releases kinetics at elevated temperature. The results of thermal analyses of immobilized flavor could be useful for further application of electrospinning process for food additives protection.

Conclusions

We showed that electrospinning technique is suitable for production of nanofibrous PVA films that contain active food ingredient. Electrospun PVA nanofibrous films with immobilized ethyl vanillin have been successfully prepared by ethanol/water mixture as a solvent. The morphology and diameters of fibers were strongly influenced by the immobilized flavor. The results of mechanical tests and FTIR analysis showed that films with immobilized ethyl vanillin were mechanically stable with clearly observed the presence of PVA and immobilized flavor in FTIR spectrum

of electrospun films. DSC analysis showed that in the range from room temperature up to 250 °C, the endothermic peaks occurred. These peaks were identified as melting points of free and immobilized ethyl vanillin and PVA. The results of this study showed that electrospinning process affected ethyl vanillin thermal properties by decreasing its melting point compared to pure flavor. In the temperature range from 250 up to 500 °C, the dominant process was thermal decomposition of PVA and ethyl vanillin.

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